

Perspective

The ARTS Connection

Role of ARTS in Apoptosis and Cancer

Sarit Larisch

Correspondence to: Sarit Larisch; Apoptosis Research Laboratory; Pathology Department; Rambam Medical Center; Haifa 31096 Israel; Tel.: 972.4.8543464; Fax: 972.4.8542877; Email: s_larisch@rambam.health.gov.il

Received 05/25/04; Accepted 06/16/04

Previously published online, as a Cell Cycle E-publication:
<http://www.landesbioscience.com/journals/cc/abstract.php?id=1021>

KEY WORDS

ARTS, apoptosis, mitochondria, caspase, IAP, cancer, tumor suppressor

ACKNOWLEDGEMENTS

I wish to thank Hermann Steller for critical reading of this manuscript and stimulating discussions. I also thank members of my lab, Yossi Gottfried, Rona Lotan, Asaf Rotem and Keren Elnekave who's work was quoted in this review. Part of this work was supported by a grant from the Israel Science Foundation.

ABSTRACT

ARTS is an unusual septin that is localized to mitochondria in living cells and promotes apoptosis by antagonizing IAPs. ARTS functions as a tumor suppressor in Acute Lymphoblastic Leukemia (ALL) and is lost in more than 70% of leukemic patients. The loss of ARTS is specific as levels of H5, a closely related non-apoptotic septin protein derived from the same gene, were unaffected. Thus, ARTS, a new member in the mitochondrial pro-apoptotic arsenal, provides a link between mitochondria, apoptosis and cancer.

ARTS belongs to the septins, a family of proteins that were originally studied for their role in cytokinesis and cell motility.¹⁻³ The ARTS protein is derived as a splice variant from the H5/PNUT12 human septin gene.⁴ Although it exhibits a high degree of sequence similarity with other family members, ARTS has several unique features. While other septins are usually localized mainly to the cytoskeleton, ARTS localizes to the mitochondria in living cells and translocates to the nucleus upon pro-apoptotic stimuli.^{4,5} The basis for the differences in cellular localization appears to be the N' terminus of ARTS, where ARTS lacks 20 amino acids found in most other septins. This change exposes a putative mitochondrial import sequence (KRFLD) which is presumably responsible for mitochondrial location of ARTS. Interestingly, another septin (M-septin) was recently reported to localize to mitochondria as well.⁶ However, M-septin contains the additional 20 amino acids at its N-terminus, suggesting that its localization in mitochondria may involve a different import mechanism than that employed by ARTS.

Evidence that ARTS plays a role in apoptosis has come both from gain and loss-of-function studies. In certain cells, high level overexpression of ARTS is sufficient to induce apoptosis (see ref. 5 and our unpublished results). More generally, expression of ARTS can promote apoptosis in response to a variety of pro-apoptotic stimuli such as, Fas, TGF- β , ara-C, etoposide, and staurosporine (STS).^{4,5,7} Conversely, downregulation of endogenous ARTS by anti-sense expression was shown to protect cells against TGF- β -induced apoptosis.⁴ In all these cases, ARTS-mediated cell killing leads to caspase activation (see below). Because ARTS is implicated in a wide variety of apoptotic paradigms, it seems to function at a central apoptotic junction where different upstream apoptotic inputs converge to mediate caspase activation.

ARTS is a mitochondrial protein that translocates from mitochondria to the nucleus during apoptosis.^{4,5} At this point, little is known about the underlying molecular mechanism. The release of ARTS from mitochondria does not require caspase activity, though this activity is required for efficient nuclear entry.⁵ The translocation of ARTS to the nucleus may not simply be the consequence of apoptosis since ARTS contains a putative nuclear import sequence (Lotan R, Larisch S, unpublished observation). Furthermore, during apoptosis ARTS becomes localized in a highly specific pattern within the nucleus, in close association with PML-NBs (promyelocytic leukemia nuclear bodies) (Fig. 1A; Lotan R, Larisch S, unpublished results). The PML-NBs are protein complexes associated with the nuclear matrix. PML constitutes the scaffold component of NBs and recruits a variety of proteins onto these domains, many of which are involved in apoptosis.^{8,9} However, the nature and reason of the association of ARTS with PML-NBs is not yet clear.

ARTS induces apoptosis through activation of caspases. In addition to activation of caspase-3, ARTS-induced-apoptosis is associated with caspase-9 activation (Fig. 1B). Whether the activation of caspase-9 by ARTS is the result of a direct effect on the apoptosome, or whether the effect is indirect remains an open question.

The main mechanism by which ARTS exerts its apoptotic activity appears to be through direct binding and inhibition of IAPs.⁵ Upon induction of apoptosis, ARTS is released from mitochondria and co-localizes with XIAP in the cytosol. Binding of ARTS to XIAP

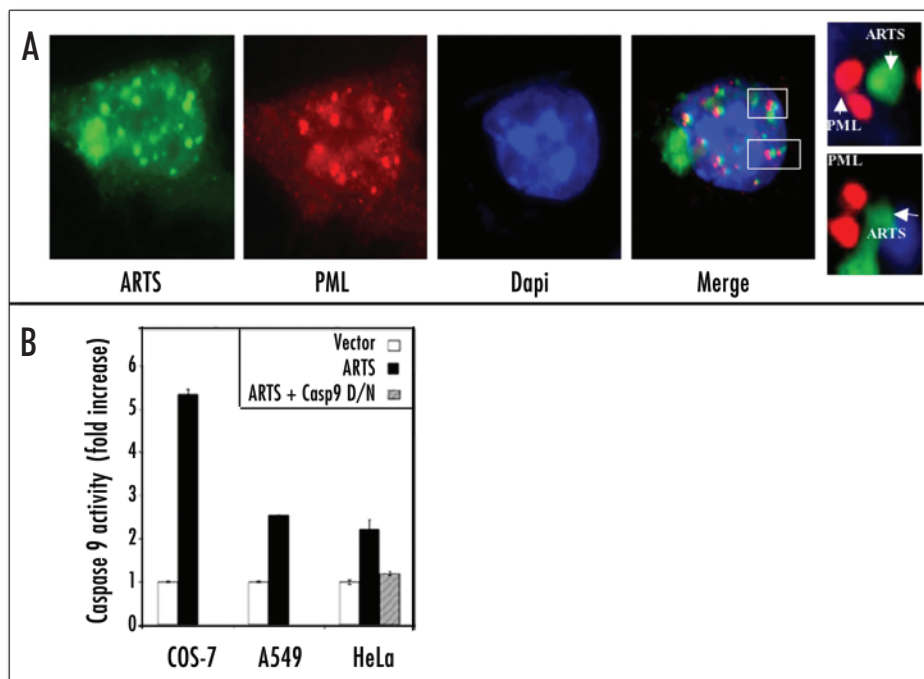


Figure 1. ARTS nuclear localization is associated with PML NBs. (A) COS-7 cells over-expressing ARTS were treated with staurosporine and subsequently stained with anti-ARTS (green) anti-PML (red) and with Dapi (blue) for nuclear localization. The merged picture shows an association of ARTS with PML NBs. Insets show an enlargement of areas in the merged picture. (B) ARTS-induced apoptosis leads to caspase-9 activation. COS-7, A549 and HeLa cell lines were transiently transfected with AU5-ARTS or AU5 empty vector. HeLa cells were also co-transfected with AU5-ARTS and caspase-9 Dominant Negative (D/N) (Cas9 D/N). Activation of caspase-9 was measured with Caspase-9 activity assay kit (R&D). The results are presented as fold increase relative to results obtained in cells transfected with control vector, which are calculated as 1. Overexpression of ARTS induces activation of caspase-9 in these cell lines, and co-transfection of ARTS and caspase-9 D/N resulted in inhibition of caspase-9 activation.

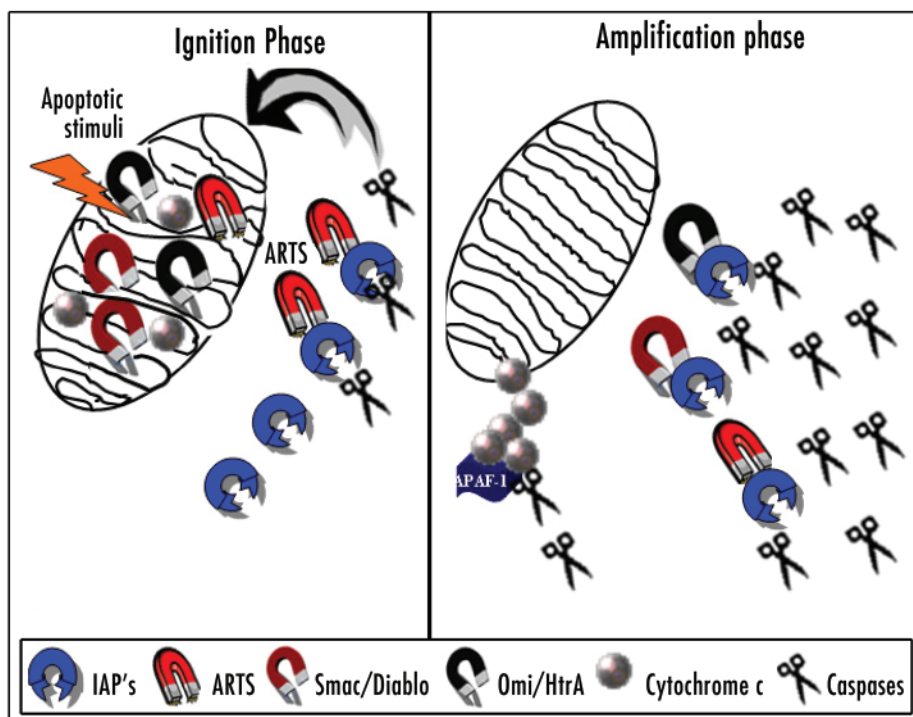


Figure 2. Two phase model for mitochondrial induced caspase activation. Proposed hypothetical model for mitochondrial induced caspase activation. The first step is an "ignition phase" in which ARTS, and possibly other caspase-independent factors, exit the mitochondria, bind to IAPs and reduce their activities. Thereby, a first sub-lethal (and possibly reversible) wave of caspase activation may be initiated. If pro-apoptotic stimuli are sufficiently strong and/or persist, this first wave of caspase activity will promote the release of cytochrome c and Smac/Diablo from mitochondria into the cytosol. As a result, an "amplification phase" will be initiated which leads to full-blown activation of caspases, cleavage and degradation of IAPs and the execution of apoptotic cell death.

is direct, as recombinant ARTS and XIAP proteins can bind to each other in vitro. ARTS binding to XIAP is specific and related to its pro-apoptotic function, as mutant forms of ARTS (and other related but non-apoptotic septins) that fail bind XIAP fail to induce apoptosis. Binding of ARTS to XIAP causes a significant reduction in XIAP levels and leads to caspase activation and cell death.⁵ Other mitochondrial pro-apoptotic proteins, such as Smac/Diablo and Omi-HtrA2, are known to act through antagonizing IAPs.¹⁰⁻¹³ How does ARTS fit within these paradigms? Does it compete with the other mitochondrial proteins for binding to IAPs or, does it complement their function?

Several unique properties of ARTS suggest the following

onists.¹⁴⁻¹⁷ Therefore, ARTS appears to bind IAPs using a novel mechanism. It is possible that binding of ARTS to XIAP requires the unique C-terminus of ARTS, a stretch of 27 amino acids not found in H5 or other septins.^{4,5} Consistent with this idea, deletion of the C-terminus of ARTS results in loss of XIAP-binding.⁵

2. ARTS appears to act upstream of Smac/Diablo and cytochrome c. The release of both Smac/Diablo and cytochrome c has been reported to require caspase activity.¹⁸⁻²² In contrast, the release of ARTS from mitochondria and its ability to bind and antagonize XIAP are not blocked by caspase inhibitors.⁵

In addition, upon apoptotic induction, ARTS levels in the cytosol peak before the release of cytochrome c.⁵ Therefore, it seems that

ARTS exits mitochondria prior to the release of both Smac/Diablo and cytochrome c.

Although both postulates await direct experimental confirmation, the available data suggest a two-phase model of caspase activation by mitochondrial factors (Fig. 2). The first step is an "ignition phase" in which ARTS, and possibly other caspase-independent factors, exit the mitochondria, bind to IAPs and reduce their activities. Thereby, a first sub-lethal (and possibly reversible) wave of caspase activation may be initiated. If pro-apoptotic stimuli are sufficiently strong and/or persist, this first wave of caspase activity will promote the release of cytochrome c and Smac/Diablo from mitochondria into the cytosol (for example, through cleavage of Bid).¹⁸ As a result, an "amplification phase" will be initiated which leads to full-blown activation of caspases, cleavage and degradation of IAPs,²³⁻²⁵ and the execution of apoptosis. The advantage of such a two-phase mechanism may be that it provides additional safety and better integration of diverse signaling pathways controlling apoptosis.

The loss of pro-apoptotic genes is one of the known mechanisms by which transformed cells obtain selective advantage through reduced susceptibility to apoptotic signals.²⁶⁻²⁸ If ARTS expression is important for the cells ability to undergo apoptosis, than we would expect cancer cells to lose ARTS expression in order to gain selective advantage. Indeed, ARTS expression is lost in lymphoblasts of more than 70% of leukemic patients.⁷ In contrast, levels of the related non-apoptotic H5 septin were unaffected in ALL patients, indicating specific selection against the pro-apoptotic function encoded by the H5/PNUTL2 gene. Interestingly, two other septins, CDCrel-1 and MSF have been found as fusion proteins with MLL (ALL1, Htrx, HRX) in de novo leukemias in children and therapy-related acute leukemia patients.^{29,30} Moreover, MSF is located on chromosome 17q25 in the vicinity of ARTS, which maps to chromosomal location 17q23.^{30,4} This region is deleted in some breast and ovarian tumors, and Kalikin et al. hypothesized that this region contains a tumor suppressor gene.³¹ Further investigation should indicate whether ARTS is this proposed tumor suppressor which contributes to the malignant transformation of hematopoietic cells, and possibly other cell types as well.

ARTS levels may critically influence IAP protein levels, both in normal and malignant cells. IAPs are over-expressed in a variety of tumors, which is thought to promote cancer cell survival,^{27,32} but the underlying molecular mechanism is still unknown. Furthermore, upregulation of IAPs is associated with resistance of tumor cells to apoptosis.^{33,34} It is possible that elevated IAPs are, at least in some tumors, the consequence of loss of ARTS. Thus, the absence of ARTS may contribute to the resistance of cancer cells to apoptosis through its direct effect on increased IAP levels.

Altogether, ARTS appears to be a central mitochondrial pro-apoptotic protein with a preferred loss in certain cancers. Therefore, strategies aimed at restoring ARTS function may provide promising new therapeutic opportunities.

References

- Field CM, Kellogg D. Septins: cytoskeletal polymers or signaling GTPases? *Trends Cell Biol* 1999; 10:387-94.
- Kartmann B, Roth D. Novel roles for mammalian septins: from vesicle trafficking to oncogenesis. *J Cell Sci* 2001; 114:839-44.
- Kinoshita M, Field CM, Coughlin ML, Straight AF, Mitchison TJ. Self-and actin-templated assembly of mammalian septins. *Dev Cell* 2002; 6:791-802.
- Larisch S, Yi Y, Lotan R, Kerner H, Eimerl S, Tony Parks W, Gottfried Y, Birkey Reffey S, de Caestecker MP, Danielpour D, Book-Melamed N, Timberg R, Duckett CS, Lechleider RJ, Steller H, Orly J, Kim SJ, Roberts AB. A novel mitochondrial septin-like protein, ARTS, mediates apoptosis dependent on its P-loop motif. *Nat Cell Biol* 2000; 2:915-21.
- Gottfried Y, Rotem A, Lotan R, Steller H, Larisch S. The mitochondrial ARTS protein promotes apoptosis through targeting XIAP. *EMBO J* 2004; 23:1627-35.
- Takahashi S, Inatome R, Yamamura H, Yanagi S. Isolation and expression of a novel mitochondrial septin that interacts with CRMP/CRAM in the developing neurones. *Genes Cells* 2003; 2:81-93.
- Elhasid R, Sahar D, Merling A, Zivony Y, Rotem A, Ben-Arush M, Izraeli S, Bercovich D, Larisch S. The mitochondrial pro-apoptotic ARTS protein is lost in the majority of Acute Lymphoblastic Leukemia patients. *Oncogene* 2004; 23:5468-75.
- Takahashi Y, Lallemand-Breitenbach V, Zhu J, de The H. PML nuclear bodies and apoptosis. *Oncogene* 2004; 23:2819-24.
- Bernardi R, Pandolfi PP. Role of PML and the PML-nuclear body in the control of programmed cell death. *Oncogene* 2003; 22:9048-57.
- Du C, Fang M, Li Y, Li L, Wang X. Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. *Cell* 2000; 102:33-42.
- Verhagen AM, Ekert PG, Pakusch M, Silke J, Connolly LM, Reid GE, Moritz RL, Simpson RJ, Vaux DL. Identification of DIABLO, a mammalian protein that promotes apoptosis by binding to and antagonizing IAP proteins. *Cell* 2000; 102:43-53.
- Verhagen AM, Silke J, Ekert PG, Pakusch M, Kaufmann H, Connolly LM, Day CL, Tikoo A, Burke R, Wrobel C, Moritz RL, Simpson RJ, Vaux DL. HtrA2 promotes cell death through its serine protease activity and its ability to antagonize inhibitor of apoptosis proteins. *J Biol Chem* 2002; 277:445-54.
- Hegde R, Srinivasula SM, Zhang X, Wassell R, Mukattash R, Cilenti L, DuBois G, Lazebnik Y, Zervos AS, Fernandes-Alnemri T, Alnemri ES. Identification of Omi/HtrA2 as a mitochondrial apoptotic serine protease that disrupts inhibitor of apoptosis protein-caspase interaction. *J Biol Chem* 2002; 277:432-8.
- Tittel J, Steller H. A comparison of programmed cell death between species. *Genome Biology* 2000; 1:reviews 31-5.
- Shi Y. A conserved tetrapeptide motif: potentiating apoptosis through IAP binding. *Cell Death Differ* 2002; 9:93-5.
- Huang Y, Park YC, Rich RL, Segal D, Myszkowski DG, Wu H. Structural basis of caspase inhibition by XIAP: differential roles of the linker versus the BIR domain. *Cell* 2001; 104:781-90.
- Vaux DL, Silke J. Mammalian mitochondrial IAP binding proteins. *Biochem Biophys Res Commun* 2003; 304:499-504.
- Li H, Zhu H, Xu CJ, Yuan J. Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. *Cell* 1998; 94:491-501.
- Bossy-Wetzel E, Green DR. Caspases induce cytochrome c release from mitochondria by activating cytosolic factors. *J Biol Chem* 1999; 274:17484-90.
- Adrain C, Creagh EM, Martin SJ. Apoptosis-associated release of Smac/DIABLO from mitochondria requires active caspases and is blocked by Bcl-2. *EMBO J* 2001; 20:6627-36.
- Guo Y, Srinivasula SM, Druilhe A, Fernandes-Alnemri T, Alnemri ES. Caspase-2 induces apoptosis by releasing proapoptotic proteins from mitochondria. *J Biol Chem* 2002; 277:13430-7.
- Hasenjager A, Gillissen B, Muller A, Normand G, Hemmati PG, Schuler M, Dorken B, Daniel PT. Smac induces cytochrome c release and apoptosis independently from Bax/Bcl-x_L in a strictly caspase-3-dependent manner in human carcinoma cells. *Oncogene* 2004; 23:4523-35.
- Ditzel M, Wilson R, Tenev T, Zachariou A, Paul A, Deas E, Meier P. Degradation of DIAP1 by the N-end rule pathway is essential for regulating apoptosis. *Nat Cell Biol* 2003; 5:467-73.
- Deveraux QL, Leo E, Stennicke HR, Welsh K, Salvesen GS, Reed JC. Cleavage of human inhibitor of apoptosis protein XIAP results in fragments with distinct specificities for caspases. *EMBO J* 1999; 18:5242-51.
- Yang Y, Fang S, Jensen JP, Weissman AM, Ashwell JD. Ubiquitin protein ligase activity of IAPs and their degradation in proteasomes in response to apoptotic stimuli. *Science* 2000; 288:874-7.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100:57-70.
- Igney FH, Krammer PH. Death and anti-death: tumor resistance to apoptosis. *Nat Rev Cancer* 2002; 2:277-88.
- Evan GI, Vousden KH. Proliferation, cell cycle and apoptosis in cancer. *Nature* 2001; 411:342-8.
- Megoni MD, Rappaport EF, Jones DH, Williams TM, Lovett BD, Kelly KM, Lerou PH, Moulton T, Budarf ML, Felix CA. t(11:22)(q23;q11.2) In acute myeloid leukemia of infant twins fuses MLL with hCDCL, a cell division cycle gene in the genomic region of deletion in DiGeorge and velocardiofacial syndromes. *Proc Natl Acad Sci USA* 1998; 95:6413-8.
- Osaka M, Rowley JD, Zeleznik-Le NJ. MSF (MLL septin-like fusion), a fusion partner gene of MLL, in a therapy-related acute myeloid leukemia with a t(11:17)(q23;q25). *Proc Natl Acad Sci USA* 1999; 96:6428-33.
- Kalikin LM, Sims HL, Petty EM. Genomic and expression analyses of alternatively spliced transcripts of the MLL septin-like fusion gene (MSF) that map to a 17q25 region of loss in breast and ovarian tumors. *Genomics* 2000; 63:165-72.
- Salvesen GS, Duckett CS. IAP proteins: blocking the road to death's door. *Nat Rev Mol Cell Biol* 2002; 3:401-10.
- Zhang J, Li Y, Shen B. Tumors acquire inhibitor of apoptosis protein (IAP)-mediated apoptosis resistance through altered specificity of cytosolic proteolysis. *Leukemia* 2002; 16:2163-5.
- Hong X, Lei L, Glas R. Inhibition of apoptosis in normal and transformed intestinal epithelial cells by cAMP through induction of inhibitor of apoptosis protein (IAP)-2. *J Exp Med* 2003; 197:1731-43.