ARTS-based anticancer therapy: taking aim at cancer stem cells

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Apoptosis related protein in TGF- β signaling pathway (ARTS/Sept4 i2) here forth referred to as ARTS, was originally found to promote apoptosis induced by TGF- β , but later was shown to promote apoptosis induced by a wide variety of apoptotic stimuli. In vitro and in vivo studies revealed that ARTS-induces apoptosis, at least in part, through direct binding and antagonizing XIAP. High levels of XIAP are found in many types of cancers and often correlate with poor prognosis. ARTS was shown to function as a tumor-suppressor protein in human patients and mouse-tumor models. In particular, Sept4/ARTS-deficient mice have increased tumor susceptibility and contain increased numbers of stem cells (SCs) and progenitor cells, apparently owing to their resistance towards apoptosis. Based on these results we propose that loss of proapoptotic ARTS may act as the 'first hit' initiating tumorigenesis in two distinct ways. First, loss of ARTS-mediated apoptosis leads to increased numbers of normal SCs. Elevated numbers of normal SCs may lead to increased cancer risk due to higher numbers of cellular targets available for transforming mutations. Second, after these SCs acquire additional transforming mutations and become cancer stem cells (CSCs), they are more likely to survive in the absence of ARTS owing to increased resistance toward apoptosis. A combination of these two mechanisms, over time, is expected to significantly increase tumor risk. Because CSCs appear to share phenotypic markers with normal SCs, targeting the signaling pathways that affect normal SC development and maintenance can serve as a useful approach towards true eradication of cancer. In this article we describe the role of ARTS in apoptosis and cancer, with focus on its potential role as a CSC marker and as a potential target for anticancer and anti-CSC therapy.

Apoptosis pathways, activators & inhibitors

Programmed cell death by apoptosis is important for regulating cell numbers and maintaining tissue homeostasis [1,2]. The main executioners of apoptosis are caspases, a family of proteases harboring a cysteine residue at their active site that preferentially cleave substrates after aspartate [3–5].

The apoptotic process is tightly controlled through the action of both activators and inhibitors of caspases [6–8]. Inhibitor of apoptosis (IAP) proteins are a major family of caspase inhibitors [9,10]. All IAP proteins contain at least one BIR domain. BIR domains can directly interact with caspases and inhibit their apoptotic activity [6,7,10,11]. Thus far, eight IAP proteins have been identified in mammals: NAIP, cIAP1, cIAP2, XIAP, MLIAP, ILP2, survivin and BRUCE/Apollon [9,12]. Some of these proteins, namely XIAP, cIAP1, cIAP2, MLIAP and ILP2, also contain a RING domain that bestows E3-ubiquitin ligase activity on these proteins [13–15]. XIAP, the best studied IAP, contains three BIR domains and can directly inhibit caspases-3, -7 and -9 [16-19]. There are two main pathways leading to caspase activation in mammalian cells [20]. The mitochondrial pathway (intrinsic pathway) and the extrinsic pathway activated through death receptors mainly in cells of the immune system. Caspase activation in the mitochondrial pathway is executed by two different modes of action (FIGURE 1). On the one hand, caspases are activated by the release of Cytochrome C (CytoC), leading to formation of a holoenzyme complex known as the apoptosome. CytoC released from the mitochondria binds to APAF-1 to activate procaspase-9 [21-23]. Second, mitochondrial factors such as SMAC/ DIABLO, Omi/Htra2 and ARTS [24-27] acting as IAP antagonists, bind to IAP proteins in the cytosol, release these caspases from their inhibition by the IAP proteins and promote their activation (FIGURE 1). XIAP also contains an E3-ubiquitin ligase activity that promotes caspase-3 ubiquitination and its subsequent proteasome-mediated degradation [14]. IAP antagonists in Drosophila as well as SMAC/DIABLO and

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Omi/Htra2 in mammalian cells use a short, conserved N-terminal sequence (AVPI) termed IAP Binding Motif (IBM) used for IAP-binding and inhibition [7,24–26,28,29]. The crystal structure of a XIAP/SMAC complex confirmed that SMAC binds the same IBM-binding grooves as the caspases [30]. The SMAC IBM is also very similar to the IBM of caspase-9 [31].

ARTS promotes apoptosis via a distinct mechanism that differs from other IAP-antagonists

ARTS is a mitochondrial protein that promotes apoptosis through binding to XIAP [27,32-34]. ARTS is derived by differential splicing from the human Septin gene *Sept4* [33,35]. Septins have been traditionally studied for their role in cytokinesis and filament forming abilities, but subsequently have been implicated in diverse functions, including determination of cell polarity, cytoskeletal reorganization, membrane dynamics, vesicle trafficking and oncogenesis [36-38]. ARTS is exceptional both in terms of its mitochondrial localization and its proapoptotic function, not shared by any other known Septin family member [39]. Moreover, ARTS promotes apoptosis via a mechanism distinct from all other known IAP antagonists. First, ARTS does not contain the canonical IBM found in most other IAP antagonists including SMAC/ DIABLO, and it binds to XIAP via a unique sequence that we term ARTS-IBM (AIBM) [40]. Second, ARTS binds to both BIR1 and BIR3 domains in XIAP [41,42]. Yet, ARTS binds to distinct sequences within BIR3, which are not bound by other IAP-antagonists such as SMAC



Figure 1.Caspase activation through the mitochondrial apoptotic pathway. In the mitochondrial pathway, caspase activation is executed through the release of proapoptotic proteins which promote activation by two different modes of action. **(A)** Release of CytoC, which leads to formation of a holoenzyme complex known as the 'apoptosome'. CytoC is released from mitochondria and binds to APAF-1 to activate procaspase 9. **(B)** Mitochondrial inhibitor of apoptosis antagonists, SMAC, OMI and ARTS, which bind and inhibit XIAP in the cytosol, thereby removing caspase inhibition. Importantly, ARTS is localized at the mitochondrial outer membrane while other proteins such as SMAC and CytoC are localized at the Intermembrane space and can be released to the cytosol only following the process of mitochondrial outermembrane permeabilization. Upon apoptotic stimuli, the translocation of ARTS from the mitochondria to the cytosol precedes the release of both CytoC and SMAC, and is required for it [43]. APAF-1: CytoC: Cytochrome C; HTRA: OMI: SMAC:

and Omi/HtrA [41]. Third, ARTS appears to initiate the mitochondrial apoptotic pathway upstream of CytoC and SMAC [43]. ARTS-XIAP complex is formed as quickly as 15-30 min after induction of apoptosis, significantly before the release of SMAC and CytoC from mitochondria, which occurs hours later [43]. Furthermore, the translocation of ARTS from the mitochondria to the cytosol is required for the on time release of CytoC and SMAC from the mitochondria, as knockdown of ARTS in HeLa cells inhibits the release of both CytoC and SMAC [43]. Finally, SMAC selectively reduces the levels of c-IAP1 and c-IAP2 but not that of XIAP [44], and SMAC-based IAP antagonists have the ability to induce degradation of cIAPs, but not XIAP [45,46]. cIAP degradation by SMAC, occurs through NF-κB activation, and TNF- α -dependent apoptosis [45]. By contrast, ARTS appears to promote apoptosis through direct binding and degradation of XIAP, and ARTS inhibits XIAP-induced NF-κB activation [42]. Collectively, it appears that ARTS functions via a distinct mechanism to promote caspase activation and tumor suppression.

Anticancer therapies targeting XIAP

High levels of IAP proteins are found in many types of cancers [47-49] and it often correlates with poor prognosis [50-52]. Therefore targeting IAP proteins presents a promising approach for developing novel anticancer drugs [48,53-55]. XIAP is considered to be the most potent inhibitor of caspases in vitro and elevated levels of this protein are found in a wide variety of human tumors [47,48,56,57]. Conversely, because mice deficient for XIAP are viable [58], the physiological function of XIAP in situ has remained unclear. However, it was later demonstrated that loss of XIAP function causes elevated caspase-3 levels and sensitizes certain primary cells towards apoptosis [14]. In addition, XIAP-mutant mice are protected against Eµ-Myc-driven lymphoma owing to increased apoptosis of premalignant lymphocytes [14,59]. Several approaches for developing anticancer drugs have focused on specifically antagonizing XIAP [60-62]. These approaches include ASOs or RNAi-based technologies selectively inhibiting expression of XIAP [49,63-66]. In addition, small molecule XIAP-antagonists were designed and tested in clinical trials [45,46,53,54,56,67-71]. Small-molecule XIAP inhibitors de-repress downstream caspases [72]. Targeting XIAP has been shown to sensitize non-small-cell lung carcinoma to y-irradiation and human ovarian and prostate cancer cells to chemotherapeutic agents *in vitro* [73–75]. In addition, inhibition of XIAP induced apoptosis and enhanced sensitivity towards chemotherapy in human prostate cancer cells [75]. Inhibition of XIAP with an antisense oligonucleotide (ASO) delayed tumor growth in a lung cancer xenograft model [76] and XIAP ASO induced apoptosis preferentially in CD34⁺38- cells in a Phase I/II study of patients with relapsed/refractory acute myeloid leukemia (AML) [77].

Designing ARTS-based anticancer therapy: targeting cancers which exhibit loss of ARTS as well as cancers overexpressing XIAP

In recent years, IAP proteins have emerged as promising targets for cancer therapy and several small-molecule IAP antagonists have been developed and are currently being evaluated in clinical trials [45,46,48]. All currently available chemical IAP-antagonists are IBM-derivatives (reaper/ SMAC mimetics) with very similar properties. These compounds initially designed to target XIAP, were found to preferentially induce degradation of cIAPs but not XIAP, thereby stimulating TNF- α production and NF- κ B activation leading to inflammatory side effects in patients.

Evidence for the physiological role of ARTS as an IAP-antagonist and a tumor suppressor protein came both from human and mouse studies. Expression of ARTS is absent in lymphoblasts of more than 70% of childhood acute lymphoblastic leukemia (ALL) and lymphoma patients [78; UNPUBLISHED DATA]. Similarly, it was recently revealed that Sept4/ARTS deficient mice develop spontaneous hematopoietic tumors [79]. This suggests that ARTS functions as a tumor suppressor protein in vivo and plays a particular important role in generation of hematopoietic cancers. The tumor suppressor function of ARTS seems to be linked to its role as an XIAP-antagonist, since these Sept4/ARTS-null mice exhibit elevated XIAP protein levels and increased resistance to cell death [79]. Importantly, the tumor and apoptosis phenotypes of Sept4/ARTS-deficient mice are all suppressed by inactivation of XIAP. These findings confirm that XIAP is a major target for ARTS-induced caspase activation and tumor suppression [79]. Altogether, ARTS specifically targets XIAP, therefore ARTS-based agonists will be distinct from all known IBM-derived IAP antagonists, which are currently developed as anticancer drugs. Moreover, since ARTS does not act through inducing TNF- α , it is unlikely to involve inflammatory side effects. In addition, ARTS-based compounds are expected to target

a wide range of cancer types by being particularly effective both against tumors exhibiting loss of ARTS, as well as for tumors expressing high levels of XIAP. These features provide a window of therapeutic opportunity for ARTS to selectively target cancer cells with minimal affects on healthy cells, which contain normal levels of both ARTS and XIAP.

Cancer stem cells: resistance to apoptosis through increased XIAP levels

Defects in apoptosis can result in the expansion of a population of transformed malignant cells. Many properties of tumors are also characteristic of stem cells (SCs) [80-83]. In particular, cancer SCs (CSCs) probably derive from normal tissue SCs or early progenitors that already possess self-renewal and unlimited proliferation potential [80,84]. Moreover, the long lifespan of normal SCs compared with short-lived differentiated progenitors or terminally differentiated cells is expected to facilitate the accumulation of genetic aberrations, which lead to cancer formation [84,85]. There is growing evidence that apoptosis plays an important physiological role in restricting the numbers of normal SCs and preventing the emergence of CSCs [86]. Evidence for resistance to apoptosis in the CD133+ fraction of glioma SCs was shown when compared with the CD133⁻ fraction [87]. Elevated levels of XIAP are associated with resistance to chemotherapy [63,88,89]. High levels of XIAP were found in CD34⁺/CD38⁻ AML SCs [90], and high levels of IAP proteins have been described in CD133⁻ population in glioblastoma [87]. In addition XIAP ASO achieves target knockdown and induced apoptosis preferentially in CD34+38cells patients with relapsed/refractory AML [77]. The relevance of the regulation of mitochondrial apoptosis in CSCs has been further demonstrated by the effective sensitization of glioblastoma-initiating cells to y-irradiationinduced apoptosis via inhibition of XIAP [91,92].

Taking into consideration that *Sept4/ARTS*null mice exhibit increased numbers of SCs with elevated levels of XIAP and increased resistance to apoptosis, it appears that loss of ARTS is at least one way in which SCs can acquire increased resistance to apoptosis.

Cancer SC markers

The concept of a small subset of SCs that could initiate tumors was extensively described in a wide range of cancers such as breast [93], brain [94], glioblastoma multiforme [95], colon [96], ovary [97] and lung cancer [98]. The pursuit to indentify CSC markers that could be targeted for therapeutic purposes has led to several surface markers suggested to be enriched in the CSC population. In many types of solid tumors, such as ovarian cancer, breast cancer, glioblastoma, colon cancer, lung cancer and rectal cancer, cell populations exhibiting CSCs have been enriched making use of single or multiple cell surface markers, such as CD133⁺ (in ovarian cancer, colon cancer, lung cancer, glioblastoma, rectal cancer), CD44⁺/CD24^{low} (in breast cancer), CD133⁺ or CD44⁺/CD117⁺ (in ovarian cancer) [84,99-105].

Importantly, Shmelkov et al. confronted the view that CD133 is a marker of CSCs in colon cancer. They show that CD133 expression is not restricted to SCs, and that both CD133+ and CD133⁻ metastatic colon cancer cells initiate tumors [106]. In addition, olfactomedin-4 has been suggested as a CSC marker for colorectal cancer cells. [107], Arachidonate 12-lipoxygenase was suggested as a marker for prostate CSCs [108] and ALDH activity was proposed to serve as a marker of CSCs in head and neck squamous cell carcinoma [109,110]. Several CSC markers have been suggested for indentifying human hepatocellular carcinoma SCs. These include EpCAM [111] and CD13, a cell surface marker specific to semiquiescent hepatocellular carcinoma SCs [112].

The cell surface markers CD34⁺/CD38⁻ were found to enrich the population of AML cells. These markers are the same markers used for the isolation of hematopoietic SCs (HSC) yet were shown to have leukemia initiating capacity when transplanted in immunodeficient (NOD/ SCID) mice [113]. Interestingly, despite examining 85 different potential melanoma CSC markers, there was no single marker that could distinguish between tumorigenic from nontumorigenic melanoma cells [114].

In human samples isolated from healthy donors, ARTS was normally expressed in CD34⁺ HSCs as well as in mature lymphocytes [78]. This suggests that the absence of ARTS in leukemic blasts was not simply the result of an incomplete cellular differentiation, but that it is rather associated with their malignant state. Interestingly, *ARTS/Sept4*-null mice contain increased numbers of HSCs and hematopoietic progenitor cells [79]. It appears that these elevated numbers of stem and progenitor cells in *ARTS/Sept4*-null mice are responsible for the increased incidence of hematopoietic tumors in these mice.

Taken into consideration that ARTS seems to be particularly important for regulating

apoptosis in SCs, it is possible that ARTS may serve as a marker to distinguish normal SCs from CSCs.

Stem cells & cancer

The first to demonstrate the ability of cells to transfer malignancy were Furth and Kahn (1937) who were able to inoculate inbred mice with cells derived from a leukemia arising in the same inbred strain [115]. They identified that only a small number, approximately 5%, of inoculations resulted in successful transplantation. Since then there has been increased interest in CSC research, improving experimental models in order to uncover the 'stemness' of CSCs. At least two main theories describing the origin of CSCs are currently debated; one suggests that CSCs represent differentiated cells that re-initiate their 'stem' features as part of, or following, malignant transformation. The other theory suggest that CSCs are mature SCs maintaining their 'stem' features while undergoing a malignant change [116]. Association between the emergence of cancer and increased numbers of SCs is seen in cases of myelodysplasia followed by the development of both ALL and AML [117,118].

However, according to the hierarchical model of CSCs, best demonstrated in AML, only a small fraction of SCs become CSCs, since during differentiation 'downstream' in the hierarchy, most cancer cells lose their tumorogenic capacity. Only CSCs that underwent irreversible epigenetic or genetic changes, and are characterized by specific markers, can transfer the disease when transplanted into immunocompromised NOD/SCID mice. Importantly, this CSC model does not apply to generation of melanoma CSCs, since several studies have shown that melanomas contain relatively large populations of tumorigenic cells. These populations of melanoma tumorigenic cells exhibited phenotypic heterogeneity that was not hierarchically organized, and underwent reversible phenotypic changes that were not associated with loss of tumorigenic potential [114,119,120].

Regardless of the CSC model suggested, it seems to be a consensus view that if SCs could be identified and their genetic abnormalities characterized, specific targeted therapy could be designed. Two recent studies involving ALL highlight the genetic and phenotypic heterogeneity of the leukemia SCs. Anderson *et al.* examined a series of pediatric ALL cases in which the *ETV6–RUNX1* gene fusion was an early or initiating genetic lesion. Using multiplexed FISH analysis they identified a progressive accumulation of up to eight genetic changes in single cells [121]. Their data suggest dynamic patterns of subclonal development that are nonlinear with a variable branching architecture. Serial transplantation in immunodeficient mice of leukemia propagating cells also showed heterogeneous genetic alternations, reflecting the diversity of subclones and their varied proliferative capacities. In parallel, Notta et al. reported very similar findings consistent with a nonlinear, branching, multiclonal model of leukemogenesis in BCR-ABL lymphoblastic leukemia samples [122]. They found that individual patient samples at diagnosis were composed of genetically diverse subclones that were related through a complex evolutionary process. These subclones also vary in their xenograft growth properties and leukemia-initiating-cell frequency [122].

Developing ARTS-based anticancer therapy: a new approach for targeting CSCs

(t12:21) Tel/AML1 is the most common chromosomal translocation in childhood ALL occurring in 25% of patients [123,124]. This translocation is often detected at birth yet, not all children bearing this translocation develop



Figure 2. Proposed model for the role of ARTS in tumorigenesis. We propose that loss of the proapoptotic ARTS protein may act as the 'first hit' initiating tumorigenesis in two distinct ways. First, loss of ARTS-mediated apoptosis leads to increased numbers of normal stem cells (pink). Elevated numbers of normal stem cells may lead to increased cancer risk owing to higher numbers of cellular targets available for transforming mutations and produce cancer stem cells (translucent). Second, after these stem cells acquire transforming mutations and become cancer stem cells, or cancer cells (blue) they are more likely to survive in the absence of ARTS owing to increased resistance toward apoptosis. A combination of these two mechanisms can explain how the loss of ARTS causes increased tumor development.

leukemia [125,126]. Expression of ARTS is lost in approximately 70% of ALL and lymphoma patients [78,79; UNPUBLISHED DATA]. Although it is in comparison a relatively small-scale study, the numbers are significant and striking. Moreover, the fact that approximately 30% of ARTS/Sept4 deficient mice develop spontaneous neoplasia as compared with no incidence of tumors seen in wt littermates [79] points to an important role that ARTS may play in initiation of leukemogenesis Sept4/ARTS-deficient mice have increased numbers of HSCs and hematopoietic progenitor cells, as indicated by both the use of markers and transplantation experiments testing for the presence of functional SCs by reconstituting the hematopoietic system of lethally irradiated recipient mice [79]. Importantly, no increase in cell proliferation was found in Sept4/ARTS-null mice, suggesting that the elevated numbers of functional SCs are owing to impaired SC apoptosis. HSPCs from Sept4/ARTS-null mice were significantly more resistant toward apoptosis than their wild-type counterparts and showed a robust increase in true clonogenic cell survival [79]. This suggests that ARTS functions as a tumor suppressor that regulates HSPC pool size by inducing apoptosis of superfluous SCs. According to this model, loss of proapoptotic ARTS function may act as the 'first hit' initiating tumorigenesis in two distinct ways. First, loss of ARTS-mediated apoptosis leads to increased numbers of normal HSPCs. Elevated numbers of normal HSPCs could lead to increased cancer risk due to the presence of the number of

cellular targets available for transforming mutations (FIGURE 2) [81-83,127]. Second, after these SCs acquire transforming mutations and become CSCs, they are more likely to survive in the absence of ARTS due to increased resistance toward apoptosis (FIGURE 2). A combination of these two proposed mechanisms, over time, is expected to significantly increase tumor risk. Consistent with this model, loss of ARTS in ALL patients was specific and related to its proapoptotic function as levels of the other, nonapoptotic splice variant of the Septin 4 gene (Sept4_i1/ H5/PNUTL2) remained intact [78]. Moreover, in mice, Sept4/ARTS function was specific for cell death of HSPCs in the hematopoietic compartment and loss of Sept4/ARTS led to long-term survival of HSPCs. Possible cooperation of loss of ARTS with other tumor-promoting events such as with (12,21) (TEL AML1), the most common translocation in childhood ALL may occur leading to leukemogeneis. Interestingly, six out of the 33 ALL patients in our study contained the t(12,21) (TEL AML1) translocation in addition to loss of ARTS [78]. Furthermore, Sept4/ ARTS null mice exhibited increased numbers of HSCs and accelerated tumor development in an Eµ-Myc background, attesting for additional functional cooperation with the *c-Myc* gene in lymphomagenesis. This collaboration is very similar to what has been described previously for overexpression of antiapoptotic proteins such as Bcl-2 [128-130].

Traditionally, cancer therapeutics have been optimized towards the majority of cells present

Executive summary

- ARTS (*Sept4 i2*) is a mitochondrial proapoptotic protein. Upon induction of apoptosis, ARTS translocates from the mitochondrial outermembrane to the cytosol where it promotes caspase activation prior to the release of Cytochrome C and SMAC/DIABLO.
- ARTS promotes caspase activation and apoptosis by directly binding and antagonizing XIAP.
- The mechanism by which ARTS antagonizes XIAP is distinct from all other known inhibitor of apoptosis (IAP) antagonists.
- ARTS was shown to function as a tumor suppressor protein in human and mouse studies.
- Apoptosis plays an important physiological role in restricting the numbers of normal stem cells (SCs) and preventing the emergence of cancer SCs.
- High levels of IAP proteins are found in several types of cancers. Thus, targeting IAP proteins presents a promising approach for developing novel anticancer drugs.
- Elevated levels of XIAP are associated with resistance to chemotherapy and are found in SCs from various cancer tissues.
- The tumor suppressor function of ARTS seems to be linked to its role as an XIAP-antagonist, since Sept4/ARTS-null mice have an increased rate of spontaneous tumors, exhibit elevated XIAP protein levels and are more resistant to cell death. Importantly, the tumor and apoptosis phenotypes of Sept4/ARTS-deficient mice are all suppressed by inactivation of XIAP. These findings confirm that XIAP is a major target for ARTS-induced caspase activation and tumor suppression.
- ARTS seems to be particularly important for regulating apoptosis in SCs. Therefore, it is possible that ARTS may serve as a marker to distinguish normal SCs from cancer SCs.
- ARTS-based compounds are expected to target a wide range of cancer types by being particularly effective against both tumors exhibiting loss of ARTS, as well as for tumors expressing high levels of XIAP. These features provide a window of therapeutic opportunity for ARTS to selectively target cancer cells with minimal affects on healthy cells, which contain normal levels of both ARTS and XIAP.

in a tumor. However, in order to completely eradicate a tumor and prevent regrowth and/ or metastases, it may be necessary to efficiently target the CSC compartment. Advances in our understanding of how CSCs escape cell death are likely to provide rational approaches to generate a new and improved class of anticancer drugs that selectively kill CSCs.

Conclusion & future perspective

The CSC hypothesis is an attractive model that explains several properties of metastatic tumors, but considerable controversy remains because of challenges to unequivocally identify CSCs. With continued advances in SC research, new sets of markers should emerge to visualize and isolate CSCs. One major feature that may distinguish normal SC from CSCs is acquired resistance towards apoptosis. In this regard, loss of ARTS expression may be a critical functional and diagnostic event common to many types of cancer. In the coming years, it will be important to critically investigate the association between cancer, apoptosis and SCs, and this will have important implications for both basic research and the clinic. For one, progress in this area will shed new insights into the origin of CSCs. Moreover, a better understanding of when and how ARTS is silenced during tumorigenesis will

Bibliography

- Meier P, Finch A, Evan G. Apoptosis in development. *Nature* 407(6805), 796–801 (2000).
- Thompson CB. Apoptosis in the pathogenesis and treatment of disease. *Science* 267(5203), 1456–1462 (1995).
- Thornberry NA, Lazebnik Y. Caspases: enemies within. *Science* 281(5381), 1312–1316 (1998).
- Nicholson DW. Caspase structure, proteolytic substrates, and function during apoptotic cell death. *Cell Death Differ*. 6(11), 1028–1042 (1999).
- Boyce M, Degterev A, Yuan J. Caspases: an ancient cellular sword of Damocles. *Cell Death Differ*. 11(1), 29–37 (2004).
- Shi Y. Mechanisms of caspase activation and inhibition during apoptosis. *Mol. Cell* 9(3), 459–470 (2002).
- Shi Y. A conserved tetrapeptide motif: potentiating apoptosis through IAP-binding. *Cell Death Differ*. 9(2), 93–95 (2002).
- Steller H. Regulation of apoptosis in Drosophila. *Cell Death Differ*. 15(7), 1132–1138 (2008).

facilitate the early identification of cancerous cells, provide new markers for the clinic, and influence the development of new therapeutic strategies. With an increasingly detailed understanding of the precise mechanism by which ARTS induces apoptosis, it should be possible to develop small-molecule mimics that provide highly efficient and specific targeting of XIAP. If 'ARTS-mimetics' are particularly effective in targeting CSCs, they would provide both a powerful research tool to investigate the role of CSCs in the origin of metastases, and ultimately provide more effective treatment of patients suffering from metastatic disease.

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Salvesen GS, Duckett CS. IAP proteins: blocking the road to death's door. *Nat. Rev. Mol. Cell Biol.* 3(6), 401–410 (2002).

A comprehensive review on inhibitor of apoptosis (IAP) proteins, the major known caspase inhibitors.

- Deveraux QL, Reed JC. IAP family proteins-suppressors of apoptosis. *Genes Dev.* 13(3), 239–252 (1999).
- Excellent review on IAPS.

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- Takahashi R, Deveraux Q, Tamm I *et al.* A single BIR domain of XIAP sufficient for inhibiting caspases. *J. Biol. Chem.* 273(14), 7787–7790 (1998).
- Liston P, Fong WG, Korneluk RG. The inhibitors of apoptosis: there is more to life than *Bcl2. Oncogene* 22(53), 8568–8580 (2003).
- Hu S, Yang X. Cellular inhibitor of apoptosis 1 and 2 are ubiquitin ligases for the apoptosis inducer SMAC /DIABLO. *J. Biol. Chem.* 278(12), 10055–10060 (2003).
- Schile AJ, Garcia-Fernandez M, Steller H. Regulation of apoptosis by XIAP ubiquitinligase activity. *Genes Dev.* 22(16), 2256–2266 (2008).

- First description of phenotypes in XIAP-mutant mice. And demonstration that XIAP acts as an E3-ubiquitin ligase to inhibit caspase.
- Yang Y, Fang S, Jensen JP, Weissman AM, Ashwell JD. Ubiquitin protein ligase activity of IAPs and their degradation in proteasomes in response to apoptotic stimuli. *Science* 288(5467), 874–877 (2000).
- Deveraux QL, Takahashi R, Salvesen GS, Reed JC. X-linked *IAP* is a direct inhibitor of cell-death proteases. *Nature* 388(6639), 300–304 (1997).
- Duckett CS, Nava VE, Gedrich RW *et al.* A conserved family of cellular genes related to the baculovirus IAP gene and encoding apoptosis inhibitors. *EMBO J.* 15(11), 2685–2694 (1996).
- Sun C, Cai M, Gunasekera AH *et al.* NMR structure and mutagenesis of the inhibitor-of-apoptosis protein XIAP. *Nature* 401(6755), 818–822 (1999).
- Sun C, Cai M, Meadows RP *et al.* NMR structure and mutagenesis of the third Bir domain of the inhibitor of apoptosis protein XIAP. *J. Biol. Chem.* 275(43), 33777–33781 (2000).

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- Degterev A, Boyce M, Yuan J. A decade of caspases. *Oncogene* 22(53), 8543–8567 (2003).
- Cain K, Bratton SB, Cohen GM. The Apaf-1 apoptosome: a large caspase-activating complex. *Biochimie* 84(2–3), 203–214 (2002).
- Schafer ZT, Kornbluth S. The apoptosome: physiological, developmental, and pathological modes of regulation. *Dev. Cell* 10(5), 549–561 (2006).
- Rodriguez J, Lazebnik Y. Caspase-9 and APAF-1 form an active holoenzyme. *Genes Dev.* 13(24), 3179–3184 (1999).
- Du C, Fang M, Li Y, Li L, Wang X. SMAC, a mitochondrial protein that promotes Cytochrome C-dependent caspase activation by eliminating IAP inhibition. *Cell* 102(1), 33–42 (2000).
- Martins LM. The serine protease Omi/HtrA2: a second mammalian protein with a Reaper-like function. *Cell Death Differ*. 9(7), 699–701 (2002).
- Verhagen AM, Ekert PG, Pakusch M *et al.* Identification of DIABLO, a mammalian protein that promotes apoptosis by binding to and antagonizing IAP proteins. *Cell* 102(1), 43–53 (2000).
- Gottfried Y, Rotem A, Lotan R, Steller H, Larisch S. The mitochondrial ARTS protein promotes apoptosis through targeting XIAP. *EMBO J.* 23(7), 1627–1635 (2004).
- Goyal L, Mccall K, Agapite J, Hartwieg E, Steller H. Induction of apoptosis by Drosophila reaper, hid and grim through inhibition of IAP function. *EMBO J.* 19(4), 589–597 (2000).
- Wang SL, Hawkins CJ, Yoo SJ, Muller HA, Hay BA. The Drosophila caspase inhibitor DIAP1 is essential for cell survival and is negatively regulated by HID. *Cell* 98(4), 453–463 (1999).
- Shiozaki EN, Chai J, Rigotti DJ et al. Mechanism of XIAP-mediated inhibition of caspase-9. Mol. Cell 11(2), 519–527 (2003).
- Srinivasula SM, Hegde R, Saleh A et al. A conserved XIAP-interaction motif in caspase-9 and SMAC /DIABLO regulates caspase activity and apoptosis. *Nature* 410(6824), 112–116 (2001).
- Larisch-Bloch S, Danielpour D, Roche NS et al. Selective loss of the transforming growth factor-β apoptotic signaling pathway in mutant NRP-154 rat prostatic epithelial cells. Cell Growth Differ. 11(1), 1–10 (2000).
- Larisch S, Yi Y, Lotan R *et al.* A novel mitochondrial septin-like protein, ARTS, mediates apoptosis dependent on its P-loop motif. *Nat. Cell Biol.* 2(12), 915–921 (2000).
- 34. Lotan R, Rotem A, Gonen H et al. Regulation

of the proapoptotic ARTS protein by ubiquitin-mediated degradation. *J. Biol. Chem.* 280(27), 25802–25810 (2005).

- Macara IG, Baldarelli R, Field CM *et al.* Mammalian septins nomenclature. *Mol. Biol. Cell* 13(12), 4111–4113 (2002).
- Longtine MS, Demarini DJ, Valencik ML et al. The septins: roles in cytokinesis and other processes. Curr. Opin Cell Biol. 8(1), 106–119 (1996).
- Hall PA, Russell SE. The pathobiology of the septin gene family. *J Pathol* 204(4), 489–505 (2004).
- Roeseler S, Sandrock K, Bartsch I, Zieger B. Septins, a novel group of GTP-binding proteins: relevance in hemostasis, neuropathology and oncogenesis. *Klin. Padiatr.* 221(3), 150–155 (2009).
- Mandel-Gutfreund Y, Kosti I, Larisch S. ARTS, the unusual septin: structural and functional aspects *Biol. Chem.* 392(8–9), 783–790 (2011).
- Edison N, Reingewertz TH, Gottfried Y et al. Peptides mimicking the unique ARTS-IAP-Binding-Motif (AIBM) can penetrate and kill cultured cancer cells. (Submitted for publication).
- Bornstein B, Gottfried Y, Edison N et al. ARTS binds to a distinct domain in XIAP–BIR3 and promotes apoptosis by a mechanism that is different from other IAP-antagonists. *Apoptosis* 2011 Sep; 16(9), 869–881 (2011).
- Garrison JB, Correa RG, Gerlic M *et al.* ARTS and Siah collaborate in a pathway for XIAP degradation. *Mol. Cell* 41(1), 107–116 (2011).
- Edison N, Zuri D, Maniv I et al. The IAPantagonist ARTS initiates caspase activation upstream of Cytochrome C and SMAC / Diablo. Cell Death Differ. (In press).
- Reveals that ARTS initiates caspase activation and apoptosis upstream of mitochondrial outermembrane permeabilization and cytochrome C release.
- Yang QH, Du C. SMAC /DIABLO selectively reduces the levels of c-IAP1 and c-IAP2 but not that of XIAP and livin in HeLa cells. J. Biol. Chem. 279(17), 16963–16970 (2004).
- Varfolomeev E, Blankenship JW, Wayson SM et al. IAP antagonists induce autoubiquitination of c-IAPS, NF-κB activation, and TNF-α-dependent apoptosis. Cell 131(4), 669–681 (2007).
- Vince JE, Wong WW, Khan N *et al.* IAP antagonists target cIAP-1 to induce TNF-α-dependent apoptosis. *Cell* 131(4), 682–693 (2007).

- Eckelman BP, Salvesen GS, Scott FL. Human inhibitor of apoptosis proteins: why XIAP is the black sheep of the family. *EMBO Rep.* 7(10), 988–994 (2006).
- Hunter AM, Lacasse EC, Korneluk RG. The inhibitors of apoptosis (IAPs) as cancer targets. *Apoptosis* 12(9), 1543–1568 (2007).
- Tamm I. AEG-35156, an antisense oligonucleotide against X-linked inhibitor of apoptosis for the potential treatment of cancer. *Curr. Opin Investig. Drugs* 9(6), 638–646 (2008).
- Augello C, Caruso L, Maggioni M *et al.* Inhibitors of apoptosis proteins (IAPs) expression and their prognostic significance in hepatocellular carcinoma. *BMC Cancer* 9, 125 (2009).
- 51. Tamm I, Richter S, Oltersdorf D *et al.* High expression levels of X-linked inhibitor of apoptosis protein and survivin correlate with poor overall survival in childhood *de novo* acute myeloid leukemia. *Clin. Cancer Res.* 10(11), 3737–3744 (2004).
- Xiang G, Wen X, Wang H, Chen K, Liu H. Expression of X-linked inhibitor of apoptosis protein in human colorectal cancer and its correlation with prognosis. *J. Surg. Oncol.* 100(8), 708–712 (2009).
- Lacasse EC, Mahoney DJ, Cheung HH, Plenchette S, Baird S, Korneluk RG. IAP-targeted therapies for cancer. *Oncogene* 27(48), 6252–6275 (2008).
- Vucic D, Fairbrother WJ. The inhibitor of apoptosis proteins as therapeutic targets in cancer. *Clin. Cancer Res.* 13(20), 5995–6000 (2007).
- Vaux DL. Inhibitor of Apoptosis (IAP) proteins as drug targets for the treatment of cancer. *F1000 Biol. Rep.* (2009).
- Oost TK, Sun C, Armstrong RC et al. Discovery of potent antagonists of the antiapoptotic protein XIAP for the treatment of cancer. J. Med. Chem. 47(18), 4417–4426 (2004).
- Tamm I, Kornblau SM, Segall H et al. Expression and prognostic significance of IAP-family genes in human cancers and myeloid leukemias. *Clin. Cancer Res.* 6(5), 1796–1803 (2000).
- Harlin H, Reffey SB, Duckett CS, Lindsten T, Thompson CB. Characterization of XIAP-deficient mice. *Mol. Cell Biol.* 21(10), 3604–3608 (2001).
- Jost PJ, Grabow S, Gray D *et al.* XIAP discriminates between type I and type II FAS-induced apoptosis. *Nature* 460(7258), 1035–1039 (2009).
- 60. Cheung HH, Lacasse EC, Korneluk RG. X-linked inhibitor of apoptosis antagonism: strategies in cancer treatment. *Clin. Cancer*

Res. 12(11 Pt 1), 3238-3242 (2006).

- Dean EJ, Ranson M, Blackhall F, Dive C. X-linked inhibitor of apoptosis protein as a therapeutic target. *Expert Opin Ther. Targets* 11(11), 1459–1471 (2007).
- Danson S, Dean E, Dive C, Ranson M. IAPs as a target for anticancer therapy. *Curr. Cancer Drug Targets* 7(8), 785–794 (2007).
- Mcmanus DC, Lefebvre CA, Cherton-Horvat G *et al.* Loss of XIAP protein expression by RNAi and antisense approaches sensitizes cancer cells to functionally diverse chemotherapeutics. *Oncogene* 23(49), 8105–8117 (2004).
- Cao C, Mu Y, Hallahan DE, Lu B. XIAP and survivin as therapeutic targets for radiation sensitization in preclinical models of lung cancer. *Oncogene* 23(42), 7047–7052 (2004).
- 65. Lima RT, Martins LM, Guimaraes JE, Sambade C, Vasconcelos MH. Specific downregulation of Bcl-2 and xIAP by RNAi enhances the effects of chemotherapeutic agents in MCF-7 human breast cancer cells. *Cancer Gene Ther.* 11(5), 309–316 (2004).
- Cummings J, Ward TH, Lacasse E *et al.* Validation of pharmacodynamic assays to evaluate the clinical efficacy of an antisense compound (AEG 35156) targeted to the X-linked inhibitor of apoptosis protein XIAP. *Br. J. Cancer* 92(3), 532–538 (2005).
- Li L, Thomas RM, Suzuki H, De Brabander JK, Wang X, Harran PG. A small molecule SMAC mimic potentiates TRAILand TNF-α-mediated cell death. *Science* 305(5689), 1471–1474 (2004).
- Sun H, Nikolovska-Coleska Z, Lu J et al. Design, synthesis, and characterization of a potent, nonpeptide, cell-permeable, bivalent SMAC mimetic that concurrently targets both the BIR2 and BIR3 domains in XIAP. J. Am. Chem. Soc. 129(49), 15279–15294 (2007).
- Tamm I, Trepel M, Cardo-Vila M et al. Peptides targeting caspase inhibitors. J. Biol. Chem. 278(16), 14401–14405 (2003).
- Zobel K, Wang L, Varfolomeev E *et al.* Design, synthesis, and biological activity of a potent SMAC mimetic that sensitizes cancer cells to apoptosis by antagonizing IAPS. *ACS Chem. Biol.* 1(8), 525–533 (2006).
- Ashwell JD. TWEAKing death. J. Cell Biol. 182(1), 15–17 (2008).
- Carter BZ, Gronda M, Wang Z et al. Small-molecule XIAP inhibitors derepress downstream effector caspases and induce apoptosis of acute myeloid leukemia cells. *Blood* 105(10), 4043–4050 (2005).
- 73. Holcik M, Yeh C, Korneluk RG, Chow T. Translational upregulation of X-linked inhibitor of apoptosis (XIAP) increases

resistance to radiation induced cell death. *Oncogene* 19(36), 4174–4177 (2000).

- Sasaki H, Sheng Y, Kotsuji F, Tsang BK. Down-regulation of X-linked inhibitor of apoptosis protein induces apoptosis in chemoresistant human ovarian cancer cells. *Cancer Res.* 60(20), 5659–5666 (2000).
- Amantana A, London CA, Iversen PL, Devi GR. X-linked inhibitor of apoptosis protein inhibition induces apoptosis and enhances chemotherapy sensitivity in human prostate cancer cells. *Mol. Cancer Ther.* 3(6), 699–707 (2004).
- 76. Hu Y, Cherton-Horvat G, Dragowska V et al. Antisense oligonucleotides targeting XIAP induce apoptosis and enhance chemotherapeutic activity against human lung cancer cells *in vitro* and *in vivo*. *Clin. Cancer Res.* 9(7), 2826–2836 (2003).
- 77. Carter BZ, Mak DH, Morris SJ et al. XIAP antisense oligonucleotide (AEG35156) achieves target knockdown and induces apoptosis preferentially in CD34⁺38⁻ cells in a Phase I/II study of patients with relapsed/ refractory AML. *Apoptosis* 16(1), 67–74 (2011).
- Elhasid R, Sahar D, Merling A *et al.* Mitochondrial proapoptotic ARTS protein is lost in the majority of acute lymphoblastic leukemia patients. *Oncogene* 23(32), 5468–5475 (2004).
- Garcia-Fernandez M, Kissel H, Brown S et al. Sept4/ARTS is required for stem cell apoptosis and tumor suppression. *Genes Dev.* 24(20), 2282–2293 (2010).

Demonstrates that ARTS plays an important role in stem-cell apoptosis and tumor suppression in the mouse.

- Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 414(6859), 105–111 (2001).
- Passegue E, Jamieson CH, Ailles LE, Weissman IL. Normal and leukemic hematopoiesis: are leukemias a stem cell disorder or a reacquisition of stem cell characteristics? *Proc. Natl Acad. Sci. USA* 100(Suppl. 1), 11842–11849 (2003).
- Clarke MF, Fuller M. Stem cells and cancer: two faces of eve. *Cell* 124(6), 1111–1115 (2006).
- Rossi DJ, Jamieson CH, Weissman IL. Stems cells and the pathways to aging and cancer. *Cell* 132(4), 681–696 (2008).

Authorative review on stem cells in cancer and disease.

 Kruyt FA, Schuringa JJ. Apoptosis and cancer stem cells: implications for apoptosis targeted therapy. *Biochem. Pharmacol.* 80(4), 423–430 (2010).

- Dalerba P, Cho RW, Clarke MF. Cancer stem cells: models and concepts. *Annu. Rev. Med.* 58, 267–284 (2007).
- Oguro H, Iwama A. Life and death in hematopoietic stem cells. *Curr. Opin Immunol.* 19(5), 503–509 (2007).
- Liu G, Yuan X, Zeng Z *et al.* Analysis of gene expression and chemoresistance of CD133⁺ cancer stem cells in glioblastoma. *Mol. Cancer* 5, 67 (2006).
- Notarbartolo M, Cervello M, Poma P, Dusonchet L, Meli M, D'Alessandro N. Expression of the IAPs in multidrug resistant tumor cells. *Oncol. Rep.* 11(1), 133–136 (2004).
- 89. Li Y, Jian Z, Xia K *et al.* XIAP is related to the chemoresistance and inhibited its expression by RNA interference sensitize pancreatic carcinoma cells to chemotherapeutics. *Pancreas* 32(3), 288–296 (2006).
- Carter BZ, Mak DH, Schober WD *et al.* Simultaneous activation of p53 and inhibition of XIAP enhance the activation of apoptosis signaling pathways in AML. *Blood* 115(2), 306–314 (2009).
- Vellanki SH, Grabrucker A, Liebau S et al. Small-molecule XIAP inhibitors enhance γ – irradiation-induced apoptosis in glioblastoma. *Neoplasia* 11(8), 743–752 (2009).
- Signore M, Ricci-Vitiani L, De Maria R. Targeting apoptosis pathways in cancer stem cells. *Cancer Lett.* (2011) (Epub ahead of print).
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl Acad. Sci. USA* 100(7), 3983–3988 (2003).
- Singh SK, Hawkins C, Clarke ID *et al.* Identification of human brain tumour initiating cells. *Nature* 432(7015), 396–401 (2004).
- Galli R, Binda E, Orfanelli U *et al.* Isolation and characterization of tumorigenic, stem-like neural precursors from human glioblastoma. *Cancer Res.* 64(19), 7011–7021 (2004).
- O'Brien CA, Pollett A, Gallinger S, Dick JE. A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. *Nature* 445(7123), 106–110 (2007).
- Stewart JM, Shaw PA, Gedye C, Bernardini MQ, Neel BG, Ailles LE. Phenotypic heterogeneity and instability of human ovarian tumor-initiating cells. *Proc. Natl Acad. Sci. USA* 108(16), 6468–6473
- Eramo A, Lotti F, Sette G *et al.* Identification and expansion of the tumorigenic lung cancer stem cell population. *Cell Death Differ.* 15(3),

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504-514 (2008).

- Wang WL, Farris AB, Lauwers GY, Deshpande V. Autoimmune pancreatitisrelated cholecystitis: a morphologically and immunologically distinctive form of lymphoplaSMAC ytic sclerosing cholecystitis. *Histopathology* 54(7), 829–836 (2009).
- 100. Cui XW, Zhao FJ, Liu J et al. Suppression of AKT1 expression by small interference RNA inhibits SGC7901 cell growth *in vitro* and *in vivo. Oncol. Rep.* 22(6), 1305–1313 (2009).
- 101. Puglisi MA, Sgambato A, Saulnier N *et al.* Isolation and characterization of CD133⁺ cell population within human primary and metastatic colon cancer. *Eur. Rev. Med. Pharmacol. Sci.* 13(Suppl. 1), 55–62 (2009).
- 102. Chen KL, Pan F, Jiang H et al. Highly enriched CD133(*)CD44 (*) stem-like cells with CD133(*)CD44(^{high}) metastatic subset in HCT116 colon cancer cells. *Clin. Exp. Metastasis* (2011) (Epub ahead of print).
- 103. Zhang S, Balch C, Chan MW *et al.* Identification and characterization of ovarian cancer-initiating cells from primary human tumors. *Cancer Res.* 68(11), 4311–4320 (2008).
- Ailles LE, Weissman IL. Cancer stem cells in solid tumors. *Curr. Opin Biotechnol.* 18(5), 460–466 (2007).
- Cho RW, Clarke MF. Recent advances in cancer stem cells. *Curr. Opin Genet. Dev.* 18(1), 48–53 (2008).
- 106. Shmelkov SV, Butler JM, Hooper AT et al. CD133 expression is not restricted to stem cells, and both CD133⁺ and CD133⁻ metastatic colon cancer cells initiate tumors. J. Clin. Invest. 118(6), 2111–2120 (2008).
- 107. Van Der Flier LG, Haegebarth A, Stange DE, Van De Wetering M, Clevers H. *OLFM4* is a robust marker for stem cells in human intestine and marks a subset of colorectal cancer cells. *Gastroenterology* 137(1), 15–17 (2009).
- 108. Yin B, Yang Y, Zhao Z et al. Arachidonate 12-lipoxygenase may serve as a potential marker and therapeutic target for prostate cancer stem cells. Int. J. Oncol. 38(4), 1041–1046
- 109. Clay MR, Tabor M, Owen JH *et al.* Single-marker identification of head and neck squamous cell carcinoma cancer stem cells with aldehyde dehydrogenase. *Head Neck* 32(9), 1195–1201

- 110. Chen YC, Chen YW, Hsu HS et al. Aldehyde dehydrogenase 1 is a putative marker for cancer stem cells in head and neck squamous cancer. Biochem. Biophys. Res. Commun. 385(3), 307–313 (2009).
- Terris B, Cavard C, Perret C. EpCAM, a new marker for cancer stem cells in hepatocellular carcinoma. J. Hepatol. 52(2), 280–281
- 112. Haraguchi N, Ishii H, Mimori K *et al.* CD13 is a therapeutic target in human liver cancer stem cells. *J. Clin. Invest.* 120(9), 3326–3339
- 113. Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat. Med.* 3(7), 730–737 (1997).
- Shackleton M, Quintana E, Fearon ER, Morrison SJ. Heterogeneity in cancer: cancer stem cells versus clonal evolution. *Cell* 138(5), 822–829 (2009).
- Thorough review describing cancer stem cell models, identification and characterization of cancer stem cells and cancer cell heterogeneity.
- Furth J, Kahn M. The transmission of leukaemia of mice with a single cell. *Am. J. Cancer* 31, 276–282 (1937).
- Bomken S, Fiser K, Heidenreich O, Vormoor J. Understanding the cancer stem cell. *Br. J. Cancer* 103(4), 439–445 (2010).
- 117. Domen J, Cheshier SH, Weissman IL. The role of apoptosis in the regulation of hematopoietic stem cells: overexpression of Bcl-2 increases both their number and repopulation potential. *J. Exp. Med.* 191(2), 253–264 (2000).
- 118. Yilmaz OH, Valdez R, Theisen BK *et al.* PTEN dependence distinguishes haematopoietic stem cells from leukaemia-initiating cells. *Nature* 441(7092), 475–482 (2006).
- 119. Quintana E, Shackleton M, Foster HR *et al.* Phenotypic heterogeneity among tumorigenic melanoma cells from patients that is reversible and not hierarchically organized. *Cancer Cell* 18(5), 510–523 (2010).
- Quintana E, Shackleton M, Sabel MS, Fullen DR, Johnson TM, Morrison SJ. Efficient tumour formation by single human melanoma cells. *Nature* 456(7222), 593–598 (2008).
- 121. Anderson K, Lutz C, van Delft FW et al. Genetic variegation of clonal architecture and propagating cells in leukaemia. Nature

469(7330), 356-361 (2011).

- Notta F, Mullighan CG, Wang JC et al. Evolution of human BCR–ABL1 lymphoblastic leukaemia-initiating cells. *Nature* 469(7330), 362–367 (2011).
- 123. Harrison CJ, Haas O, Harbott J et al. Detection of prognostically relevant genetic abnormalities in childhood B-cell precursor acute lymphoblastic leukaemia: recommendations from the Biology and Diagnosis Committee of the International Berlin-Frankfurt-Munster study group. Br. J. Haematol. 151(2), 132–142 (2010).
- 124. Shurtleff SA, Buijs A, Behm FG *et al.* TEL/ AML1 fusion resulting from a cryptic t(12;21) is the most common genetic lesion in pediatric ALL and defines a subgroup of patients with an excellent prognosis. *Leukemia* 9(12), 1985–1989 (1995).
- 125. Mori H, Colman SM, Xiao Z et al. Chromosome translocations and covert leukemic clones are generated during normal fetal development. Proc. Natl Acad. Sci. USA 99(12), 8242–8247 (2002).
- 126. Lausten-Thomsen U, Madsen HO, Vestergaard TR, Hjalgrim H, Nersting J, Schmiegelow K. Prevalence of t(12;21) [ETV6–RUNX1]-positive cells in healthy neonates. *Blood* 117(1), 186–189 (2011).
- 127. Tan BT, Park CY, Ailles LE, Weissman IL. The cancer stem cell hypothesis: a work in progress. *Lab. Invest.* 86(12), 1203–1207 (2006).
- Bissonnette RP, Echeverri F, Mahboubi A, Green DR. Apoptotic cell death induced by c-Myc is inhibited by Bcl-2. *Nature* 359(6395), 552–554 (1992).
- 129. Strasser A, Harris AW, Bath ML, Cory S. Novel primitive lymphoid tumours induced in transgenic mice by cooperation between Myc and Bcl-2. *Nature* 348(6299), 331–333 (1990).
- Pelengaris S, Khan M, Evan G. c-Myc: more than just a matter of life and death. *Nat. Rev. Cancer* 2(10), 764–776 (2002).